

Comparative Expression Analysis Reveals Relationships between SPINK1, TUBB3, and EZH2, and Prostate Cancer Molecular Biomarkers in the Cancer Genome Atlas (TCGA) Data

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Abstract

Background: Despite the recent discovery of molecular subtypes in prostate cancer (PCa) expressing or not gene fusions involving E26 Transformation-Specific (ETS) transcription factors, including ERG (for v-ets avian erythroblastosis virus E26 oncogene homolog), little is known on molecular alterations associated, and cooperative events at play during initiation and progression of PCa.

Objective and methods: Using RNA-Seq data from The Cancer Genome Atlas (TCGA) collection of surgically managed primary prostate adenocarcinomas, we investigated the relations between gene expression of the candidate prognostic markers *SPINK1*, *TUBB3* (class III beta-tubulin), *EZH2*, and known PCa molecular markers. 484 cases were included in the analysis.

Results: Clustering analysis consistently showed TUBB3 associating with EZH2, and SPINK1 with PTEN and TFF3, but not with ERG, ETV1, ETV1, ETV1, ETV2, ETV3, ETV3, ETV3, ETV3, ETV3, ETV3, ETV4, ETV4,

Conclusions: Despite substantial heterogeneity among the PCa cases, the current study suggests that significant associations and overlaps exist between PCa molecular alterations and expression of candidate PCa prognostic markers. A better understanding of these alterations and their cooperative role should help refine PCa subtypes, identify aggressive subgroups among those, and improve PCa management and therapy response.

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Introduction

Prostate cancer (PCa), as many other cancers, is characterized by extensive clinical and molecular heterogeneity [1]. Over the past 10 years, with the advent of high throughout methods, our understanding of the PCa genome has significantly changed, while revealing considerable intertumor (between tumors of the same type), and intra-tumor (within tumors, different subclones) heterogeneities [2-8]. Based on the molecular alterations identified, different molecular PCa subclasses or subtypes have emerged with the attempt to correlate those PCa subtypes to clinical features, disease progression and response to therapy. Approximately 50% of PCAs harbor a gene fusion between ERG, an ETS transcription factor, and an androgen-regulated gene (TMPRSS2 ~ 90%, SLC45A3, NDRG1, HERPUD1, or others <10%) [9,10]. ERG expression is routinely used as a surrogate marker of these alterations [11]. Trefoil factor 3 (TFF3) represents a highly specific

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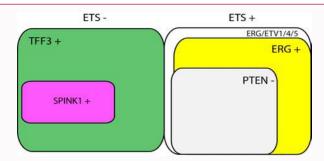


Figure 1: Schematic representation of the PCa molecular subtypes deriving from previous studies. Tumors without or with ETS fusions are shown on the left and right, respectively. In the ETS positive tumors, the vast majority harbor genomic rearrangements leading to ERG overexpression, thus representing an ERG positive PCa subtype. PTEN loss is commonly found in this subtype. ETS negative PCas generally express TFF3 as a molecular cancer biomarker, and a subgroup of these PCas express SPINK1. This subgroup may reflect a subset of disease with more aggressive behavior.

molecular biomarker of cancer in the prostate. Detectable in 40% to 60% of PCa cases [12,13], it appeared to be inversely correlated with ERG expression in most instances [14] (Figure 1). Inactivation of tumor suppressor phosphatase and tensin homolog (PTEN) is also commonly found in PCa and could be associated with cancer progression [15,16]. Several studies have found that PTEN alterations are enriched in ERG-over expressing PCa. Moreover, ERG overexpression and alterations of PTEN could cooperate, leading to more aggressive disease [17,18]. Despite these significant advances, it remains a challenge to link this molecular classification with clinical features to improve prognostic estimation, and treatment decisions in routine clinical practice [12,13,19-27].

SPINK1 (previously referred to as TATI, or tumor-associated trypsin inhibitor) is expressed in various diseases including cancer [28]. Tomlins et al. [29] identified SPINK1 as a candidate marker for a group of PCa devoid of ETS gene fusions associated with aggressive disease features and adverse outcomes. In other studies, a correlation between SPINK1 expression and adverse prognosis was not observed [25,30,31]. Nevertheless, in a recent survey, the prognostic value of SPINK1 was confirmed in a well-annotated cohort [14]. We proposed that SPINK1 overexpression emerges from a subgroup of PCa with ERG negative/TFF3 (trefoil factor 3) positive pattern [14] (Figure 1).

It is to note that various experimental studies have shown that ERG, TFF3 and SPINK1 are all associated with increased cell motility and/or invasive behavior in PCa models supporting their role in PCa progression [29,32,33].

Elevated β III-tubulin (encoded from TUBB3 gene) expression was previously identified as significantly associated with tumor aggressiveness in PCa patients with presumed localized disease [34]. In this study, β III-tubulin expression was found to be an independent marker of disease recurrence after local treatment. Recently, Tsourlakis and colleagues examined a large European cohort and confirmed this finding [35]. Additionally, increased β III-tubulin expression is associated with the emergence of Castrate Resistant PCa (CRPC) [36,37], and with lower survival for patients receiving docetaxel-based chemotherapy [34].

The polycomb group protein enhancer of zeste homolog 2 (*EZH2*) is known to be increased in metastatic PCas. Clinically localized PCa that express elevated levels of EZH2 show a poorer prognosis [38], suggesting that EZH2 has a potential role in disease progression

and patient prognosis [39-41]. Moreover *EZH2* expression is found elevated in Neuroendocrine PCa, a higly aggressive form of human PCa [42,43]. In the era of precision medicine [44,45], these findings underscore the potential utility of decrypting the relationships at play between *SPINK1*, *TUBB3*, *EZH2* expression and other PCa molecular alterations in order to improve our definition of molecular PCa subclasses and find best therapeutic solutions for the management of patients.

In the present work, we examined publically available gene expression data from primary PCa cases of The Cancer Genome Atlas (TCGA), and studied relationships between gene expression of *TUBB3*, *SPINK1*, *EZH2* and expression of other known molecular PCa biomarkers including *ERG*, *ETV1*, *PTEN*, *CHD1*, *TFF3*, *MYC*, *RB1*, *MYC*, *AURKA*, and *SPOP* [2,3,6,7,42, 46-49].

Methods

RNA-Seq gene expression analysis, clustering and statistical analysis

Human samples analyzed consisted of primary of prostate adenocarcinomas from The Cancer Genome Atlas (TCGA) project collection (http://cancergenome.nih.gov/). TCGA RNA-Seq expression data and sample information were accessed before June 2016 from cBioPortal [50] and the TCGA public access data (http://tcga-data.nci.nih.gov/). Only cases with available expression data, and analyzed for mutational landscape were considered. The cohort consisted of men surgically managed for localized or locally advanced disease. Of note, about 16% (41 of 260 cases; NA for the remaining cases) also received adjuvant treatments consisting of hormone therapy, radiotherapy, or a combination of those. Available patient cohort characteristics are shown in Table 1 (n=484).

To explore expression levels and associations of the different genes, gene expression levels (RSEM) were subjected to correlation and unsupervised clustering analyses using Cluster and TreeView softwares after transforming the RSEM into Log, (RSEM+1). Genes

 Table 1: Clinico-pathological characteristics of the TCGA studied cohort (n=484).

Age , year	
mean	61
median (range)	61 (41-78)
Pre-Operative PSA, ng/mL*	7.4 (1.6-87)
Pathological Gleason Score, no. (%)	
6	44 (9.1)
7 (3+4)	144 (30)
7 (4+3)	98 (20)
8	63 (13)
9	132 (27.3)
10	3 (0.6)
Pathological Stage, no. (%)	
pT2	183 (38.3)
рТ3а	155 (32.4)
pT3b	130 (27.2)
pT4	10 (2.1)
Lymph nodes, no. (%)	
negative	318 (80.5)
positive	77 (19.5)

*from 187 cases

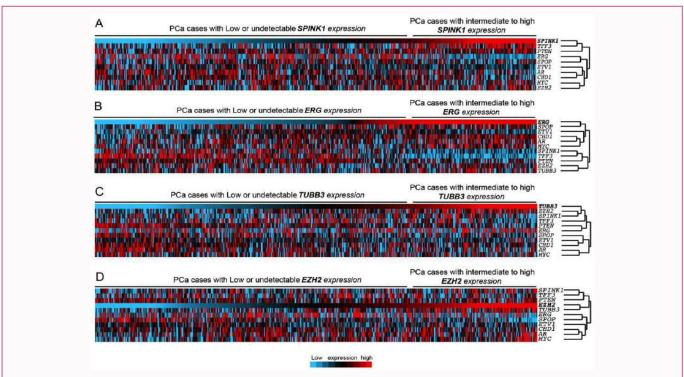


Figure 2: Hierarchical cluster analysis and heatmap generated using SPINK1, ERG, TFF3, ETV1, AR, CHD1, PTEN, SPOP, ERG, and EZH2 expressions across TCGA prostate adenocarcinomas (n=484). In the heatmap, each column represents a different case, and each row represents a marker. blue to red: lowest to highest expression. PCa cases are ordered with respect to expression of SPINK1 (A), ERG (B), TUBB3 (C), or EZH2 (D).

analyzed included *ERG*, *ETV1*, *PTEN*, *CHD1*, *TFF3*, *MYC*, *SPOP*, *SPINK1*, *EZH2*, *TUBB3*, *RB1*, and *AURKA*. Pearson's coefficient was determined to assess correlations between expression levels. For differential expression analysis, an unpaired t test or a non-parametric Mann-Whitney U test was applied as appropriate. All p values were two-sided and values of p <0.05 were considered statistically significant.

Results

In the group of ETS negative PCa, SPINK1 expression is positively correlated with TFF3 and PTEN expression levels

Expression data were retrieved from TCGA collection, tumor samples ordered by *SPINK1* expression, and clustering analysis was performed for gene expression of PCa molecular biomarkers including *ERG*, *TFF3*, *ETV1*, *AR*, *CHD1*, *PTEN*, *SPOP*, *ERG*, and *EZH2* (Figure 2a). Expectedly, *SPINK1* expression clustered with *TFF3* expression, and seems inversely correlated with *ERG* and *ETV1* expression. *SPINK1* and *TFF3* also clustered with *PTEN* expression. *PTEN* expression appeared to be especially elevated in a number of cases expressing high levels of *SPINK1*. By contrast, it was reduced in the cases expressing high levels of *ERG*, as also evidenced by an additional heatmap with tumor classified with respect to *ERG* expression (Figure 2b).

To substantiate these results, we computed correlation scores between SPINK1 expression and each molecular marker (Table 2). SPINK1 and TFF3 were significantly correlated (r=0.36, p <0.0001) and both inversely correlated with ERG, CHD1, and AR (Table 2). Moreover, ERG negatively correlated with PTEN (r=-0.2911; p <0.0001). This likely reflects an enrichment of PTEN deletion in ERG+PCa subtype as described previously [17,51,52].

A subset of tumors expressing SPINK1 concomitantly expresses high levels of EZH2

Interestingly, we also noted in the heatmap a relative enrichment of *EZH2* expression in tumors expressing high levels of *SPINK1* (Figure 2a), and based on the correlation analysis, EZH2 expression was positively correlated with *SPINK1* expression (r=0.1554, p=0.0006). This data suggests that at least a subset of *SPINK1* expressing tumors also expresses high levels of *EZH2*. Importantly, there was however no correlation between *EZH2* and *TFF3* (Table 2), neither with ERG. We previously described the presence of *SPINK1* expression characterizes an aggressive subtype in the group of ERG-/TFF3+PCa tumors [14]. Our observation here suggests that *EZH2* overexpression preferentially arises from ERG-/TFF3+/SPINK1+PCas rather than in ERG-/TFF3+/SPINK1-PCas, which could coincide with more aggressive forms of the disease.

A subset of tumors expressing *TUBB3* concomitantly expresses high levels of *EZH2*

We then investigated how these markers could cluster with TUBB3 (encoding for Class III β -tubulin), another candidate marker for aggressive PCa disease [34], also assumed to be an early marker for reduced AR signaling [36] and NE differentiation [37,53]. A Heatmap of the same set of genes in tumors classified by TUBB3 expression revealed a marked enrichment of EZH2 in tumors overexpressing TUBB3 (Figure 2c), further highlighted by a positive correlation coefficient (r=0.32; p <0001; Table 2). The analysis also revealed TUBB3 and PTEN expression patterns inversely correlated (r=-0.3159; p <0001), with AR and SPOP also following this trend (r=-0.26 and -0.20, respectively; p<0001). Intriguingly, SPINK1 expression did not appear to be associated with TUBB3 expression. SPINK1 expression was correlated with PTEN expression, when

Table 2:

Correlation with SPINK1 expression			Correlation with TFF3 expression						
Gene Pearson r 95% CI P Valu				Gene Pearson r 95% CI P Value					
TFF3	0.3659	0.2861 to 0.4407	<0.0001	ERG	-0.6027	-0.6566 to -0.5428	<0.0001		
ERG	-0.3166	-0.3947 to -0.2341	<0.0001	CHD1	-0.4702	-0.5369 to -0.3977	<0.0001		
CHD1	-0.2485	-0.3304 to -0.1630	<0.0001	SPINK1	0,3659	0.2861 to 0.4407	<0.0001		
PTEN	0.1925	0.1052 to 0.2769	<0.0001	AR	-0.2216	-0.3048 to -0.1352	<0.0001		
ETV1	-0.1587	-0.2444 to -0.07052	0.0005	ETV1	-0.2144	-0.2979 to -0.1277	<0.0001		
AR	-0.1561	-0.2419 to -0.06793	0.0006	PTEN	0.156	0.06782 to 0.2418	0.0006		
EZH2	0.1554	0.06714 to 0.2412	0.0006	SPOP	-0.1091	-0.1963 to -0.02012	0.0164		
SPOP	-0.06134	-0.1497 to 0.02796	0.1779	MYC	-0.1399	-0.2263 to -0.05141	0.002		
TUBB3	0.05186	-0.03746 to 0.1404	0.2548	TUBB3	-0,04352	-0.1322 to 0.04581	0.3393		
MYC	0.0372	-0.05212 to 0.1259	0.4141	EZH2	0.01475	-0.07449 to 0.1038	0.7461		
	Correlation with ERG expression				Correlation with PTEN expression				
Gene	Pearson r	95% CI	P Value	Gene	Pearson r	95% CI	P Value		
TFF3	-0.6027	-0.6566 to -0.5428	< 0.0001	TUBB3	-0.3159	-0.3939 to -0.2333	<0.0001		
SPINK1	-0.3166	-0.3947 to -0.2341	< 0.0001	ERG	-0.2911	-0.3706 to -0.2073	<0.0001		
CHD1	0.3161	0.2335 to 0.3941	< 0.0001	SPOP	0.2027	0.1156 to 0.2867	<0.000′		
PTEN	-0.2911	-0.3706 to -0.2073	< 0.0001	SPINK1	0.1925	0.1052 to 0.2769	<0.0001		
SPOP	0.2367	0.1507 to 0.3191	< 0.0001	TFF3	0.156	0.06782 to 0.2418	0.0006		
ETV1	-0.1408	-0.2271 to -0.05228	0.0019	EZH2	-0.1378	-0.2242 to -0.04924	0.0024		
MYC	0.112	0.02308 to 0.1992	0.0137	AR	0.13	0.04133 to 0.2166	0.0042		
TUBB3	0.1118	0.02288 to 0.1990	0.0139	MYC	0.1197	0.03089 to 0.2067	0.0084		
AR	0.06206	-0.02724 to 0.1504	0.1728	ETV1	0.04871	-0.04061 to 0.1373	0.2848		
EZH2	-0.05982	-0.1482 to 0.02949	0.1889	CHD1	0.02522	-0.06408 to 0.1141	0.58		
	Correlation	with TUBB3 expression		Correlation with EZH2 expression					
Gene	Pearson r	95% CI	P Value	Gene	Pearson r	95% CI	P Value		
EZH2	0.3246	0.2424 to 0.4021	< 0.0001	SPOP	-0,455	-0.5229 to -0.3813	< 0.000		
PTEN	-0.3159	-0.3939 to -0.2333	< 0.0001	TUBB3	0.3246	0.2424 to 0.4021	< 0.000		
AR	-0.2655	-0.3464 to -0.1806	< 0.0001	MYC	0.2668	0.1819 to 0.3476	< 0.000		
SPOP	-0.2031	-0.2870 to -0.1160	< 0.0001	SPINK1	0.1554	0.06714 to 0.2412	0.0006		
CHD1	-0.1719	-0.2571 to -0.08405	0.0001	AR	0.1381	0.04956 to 0.2245	0.0023		
ERG	0.1118	0.02288 to 0.1990	0.0139	PTEN	-0.1378	-0.2242 to -0.04924	0.0024		
MYC	-0.07149	-0.1596 to 0.01778	0.1163	CHD1	-0.08513	-0.1730 to 0.004048	0.0613		
ETV1	-0.05646	-0.1449 to 0.03286	0.215	ERG	-0.05982	-0.1482 to 0.02949	0.1889		
SPINK1	0.05186	-0.03746 to 0.1404	0.2548	ETV1	-0.03071	-0.1195 to 0.05860	0.5002		
TFF3	-0.04352	-0.1322 to 0.04581	0.3393	TFF3	0.01475	-0.07449 to 0.1038	0.7461		

 $\label{eq:Red:significant} \textbf{Red: significant positive correlation; Blue: significant negative correlation}$

TUBB3 expression anticorrelated with PTEN expression. Moreover, TUBB3 expression seemed to be more closely associated with EZH2 than SPINK1. This indicates that SPINK1 or TUBB3 expressions may be mutually exclusive under some circumstances; thus representing two distinct subsets of disease. Another possibility might be that a large proportion of TUBB3 expressing tumors is confined to the ERG+ / PTENlow PCa subtype, that is negative for SPINK1, while a smaller fraction is linked to ETS-TFF3+SPINK1 PCa subtype, and both can exhibit an enriched expression of EZH2.

EZH2 expression is associated with AURKA expression and SPOP alterations

We then generated a heatmap classifying tumors with respect

to EZH2 expression (Figure 2d), in conjunction with correlation analysis. This denoted a striking positive correlation between EZH2 and AURKA levels (r=0.7178; p <0.0001), and an inverse correlation between EZH2 and SPOP levels (r=-0.455; p <0001; Table 2). The connection between EZH2 expression and downregulated levels of SPOP is intriguing, especially considering recent work by Barbieri and colleagues who identified inactivating mutation in SPOP gene as the most common point mutation in PCa [6]. Further work by this group revealed that this mutation occurs predominantly in the group of ERG rearranged PCa tumors [6], and is concomitant with deletions at 5q21 CDH1 locus [47]. We then sought to determine whether mutation in SPOP gene, is associated with varying levels of SPINK1, EZH2 and TUBB3 (Figure 3). When considering all patients,

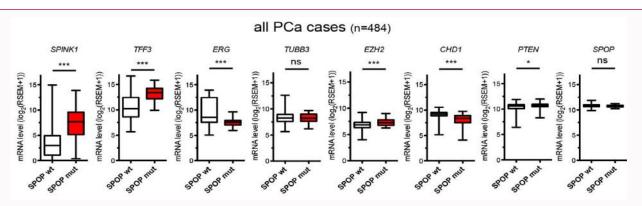


Figure 3: The effect of SPOP mutation on expression of *SPINK1*, *TFF3*, *ERG*, *TUBB3*, *EZH2*, *CHD1*, *PTEN*, and *SPOP* accross 484 PCa samples. Box plots showing the Median, 25th to 75th percentiles. Lower and upper bars correspond to the minimum and maximum values, respectively.

Table 3:

Correlation with TUBB3 expression			Correlation with EZH2 expression					
Gene	Pearson r	95% CI	P Value	Gene	Pearson r	95% CI	P Value	
SYP	0.3308	0.2490 to 0.4079	<0.0001	AURKA	0.7178	0.6716 to 0.7584	<0.0001	
EZH2	0.3246	0.2424 to 0.4021	<0.0001	TUBB3	0.3246	0.2424 to 0.4021	<0.0001	
RB1	-0.3256	-0.4030 to -0.2435	<0.0001	ENO2	-0.2655	-0.3464 to -0.1806	<0.0001	
REST	-0.3178	-0.3957 to -0.2353	<0.0001	SRRM4	-0.2319	-0.3146 to -0.1458	<0.0001	
AR	-0.2655	-0.3464 to -0.1806	<0.0001	CHGA	-0.1952	-0.2795 to -0.1079	<0.0001	
AURKA	0.2317	0.1455 to 0.3143	<0.0001	CHGB	-0.142	-0.2283 to -0.05353	0.0017	
ENO2	0.2263	0.1399 to 0.3092	<0.0001	AR	0.1381	0.04956 to 0.2245	0.0023	
CHGB	0.1169	0.02808 to 0.2040	0.01	SYP	0.06937	-0.01990 to 0.1576	0.1275	
SRRM4	0.05351	-0.03581 to 0.1420	0.24	RB1	-0.06689	-0.1551 to 0.02240	0.1417	
MYCN	-0.004543	-0.09366 to 0.08464	0.9206	REST	0.0573	-0.03201 to 0.1457	0.2083	
CHGA	0.00007283	-0.08908 to 0.08922	0.9987	MYCN	0.01582	-0.07344 to 0.1048	0.7285	
	Correlation with SPINK1 expression				Correlation with TFF3 expression			
Gene	Pearson r	95% CI	P Value	Gene	Pearson r	95% CI	P Value	
SYP	0.2342	0.1481 to 0.3167	< 0.0001	SYP	0.399	0.3213 to 0.4714	< 0.0001	
RB1	-0.2053	-0.2892 to -0.1183	< 0.0001	REST	-0.2942	-0.3735 to -0.2105	< 0.0001	
REST	-0.1742	-0.2593 to -0.08642	0.0001	RB1	-0.2566	-0.3380 to -0.1713	< 0.0001	
EZH2	0.1554	0.06714 to 0.2412	0.0006	CHGB	-0.236	-0.3184 to -0.1500	< 0.0001	
AR	-0.1561	-0.2419 to -0.06793	0.0006	AR	-0.2216	-0.3048 to -0.1352	< 0.0001	
AURKA	0.06712	-0.02217 to 0.1553	0.1404	SRRM4	-0.2085	-0.2923 to -0.1216	< 0.0001	
SRRM4	-0.06132	-0.1497 to 0.02798	0.178	ENO2	-0.1875	-0.2721 to -0.09997	< 0.0001	
CHGA	0.03842	-0.05091 to 0.1271	0.399	CHGA	-0.173	-0.2582 to -0.08516	0.0001	
CHGB	-0.01088	-0.09994 to 0.07834	0.8112	MYCN	-0.1562	-0.2420 to -0.06800	0.0006	
ENO2	0.007363	-0.08184 to 0.09645	0.8716	AURKA	-0.03589	-0.1246 to 0.05344	0.4309	
MYCN	-0.001008	-0.09015 to 0.08815	0.9824	EZH2	0.01475	-0.07449 to 0.1038	0.7461	

SPOP mutant cases had elevated expression of *EZH2*, *SPINK1* and *TFF3*, but reduction in expression of *ERG* and *CHD1*. There were no significant changes noted for *TUBB3* and *SPOP* expression levels.

SPINK1, TUBB3 and EZH2 are associated with various NE features

Previous studies have reported associations of TUBB3 and EZH2 overexpression with aggressive features in localized PCa, or NE features in castrate resistant CRPC tumors [34-36,42]. We asked weather SPINK1, TUBB3, EZH2 could also associate with NE features in this cohort of locally managed PCa tumors. Heatmaps were

generated with respect to *TUBB3*, *EZH2*, or *SPINK1* expression, and their distribution was studied as above among a panel of NE markers, putative drivers and suppressors of NE phenotype, (NE suppressors (*RB1*, *REST*, *AR*); NE drivers (*AURKA*, *SRRM4*, *MYCN*); *NE* markers (*SYP* (synpatophysin), *ENO2* (NSE), *CHGA* (chromogranin A), *CHGB* (chromogranin B)) (Figure 4). Correlation coefficients were determined as above to assess associations between variables. Notably, *EZH2* highly correlated with *AURKA* gene expression, but with the exception of *TUBB3*, showed no or negative correlation with other NE components. By contrast, tumors with high expression of *TUBB3*, and *SPINK1* to a lesser extent, more frequently exhibited NE

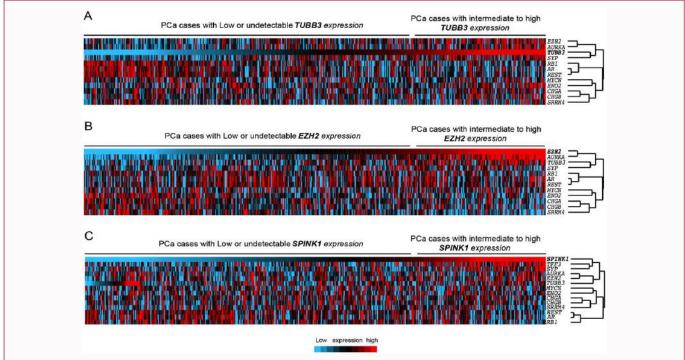


Figure 4: Hierarchical cluster analysis and heatmap generated using expression levels of SPINK1, TUBB3, EZH2, and NE components AR, SYP, RB1, CHGA, CHGB, ENO2, MYCN, AURKA, REST, SRRM4 across TCGA prostate adenocarcinomas (n=484). In the heatmap, each column represents a different case, and each row represents a marker. blue to red: lowest to highest expression. PCa cases are ordered with respect to expression of TUBB3 (A), EZH2 (B), or SPINK1 (C).

features, as judged by anti-correlations with *REST*, *AR* and *RB1*, and positive correlation with *SYP* (Table 3).

Discussion

We previously proposed SPINK1 and β III-tubulin expressions as independent prognosticators of disease recurrence in PCa patients primarily managed by prostatectomy [14,34]. Together with EZH2, another potential marker of PCa aggressiveness, these genes may be directly involved in progression, metastatic spread and/or therapy resistance of PCa. One interesting open question regarding these genes is to what extent these genes cooperate or overlap with other known molecular alterations recently characterized in PCa and defining PCa subclasses [1-3,6,7,42,46-49, 54].

In this work, by exploring RNAseq data from the TCGA prostate adenocarciomas, we confirmed on a large series of primary PCas that SPINK1 positive tumors represent a molecular subgroup of PCa tumors strongly associated with TFF3 expression, and correlating negatively with ERG expression. We found that these tumors express PTEN more often, but less CHD1 or AR. Importantly, a subset of those cases seem to overexpress EZH2. Tumors highly expressing TUBB3 also frequently exhibited higher expression of EZH2. Correlation analyses also revealed that EZH2 expression was positively associated with AURKA expression, an oncogenic target in cancer, while it was negatively associated with SPOP expression, a new putative tumor suppressor in PCa that is frequently mutated [1,6,23]. It is tempting to speculate that a molecular link exists between SPOP alterations, EZH2 and AURKA expression. In line with this possibility, our preliminary data already indicate, that in the group of SPINK1 high expressing PCa cases, SPOP mutants displayed higher expression of EZH2 and AURKA compared to SPOP wild-type (data not shown).

Altogether these findings should help refine PCa molecular subtypes, and identify subgroups of aggressive PCa. A working model

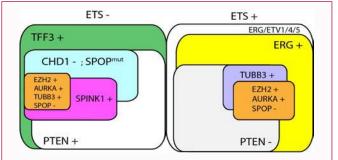


Figure 5: Schematic representation of the PCa molecular subgroups deriving from this study and from previous studies. Tumors without or with ETS fusions are shown on the left and right, respectively. In the ERG positive PCa subtype, PTEN loss is a commonly found, while it is relatively rare in ETS negative (TFF3+) subtype in which reduction of CHD1 and inactivating mutations in SPOP become more common features. In each PCa subtype, other alterations such as TUBB3, SPINK1, or EZH2 overexpression working in parallel or together (likely in association with additional related events such as AURKA upregulation and SPOP downregulation) may characterize subsets of disease with more aggressive behavior, resistant to therapies, or being able to proliferate or progress more rapidly to metastatic disease. Other important alterations, that are not shown here, are likely involved in initiation, progression, or differentiation of the disease. This includes, but not only, MYC amplification, ETV1 amplification/overexpression; mutations in TP53, CTNNB1, ATM, BRCA2, or FOXA1; deletion or reduction of NKX3.1, RB1. AR. REST.

of the different subgroups is presented in Figure 5.

It remains unclear however weather *SPOP* alterations influences the group of *TUBB3* high PCa. Aside from the apparent relations between *TUBB3*, *EZH2* and *AURKA*, *SPOP* expression was only slightly reduced in *TUBB3* high PCa, and we did not find relationship between *TUBB3* expression and *SPOP* mutation status. We posit that in the groups of *SPINK1* high or *TUBB3* high PCa tumors, also characterized by high *vs.* low expression of PTEN, respectively, a

subset of cases express significant levels of EZH2 accompanied by substantial AURKA expression which could evoke more aggressive features. A thorough assessment of such hypotheses will require further investigations on independent cohorts, and validation by various techniques including Fluorescence in Situ Hybridization (FISH) and immunohistochemistry-based approaches. Our data investigating NE features in this series indicates that EZH2 is not directly associated with NE differentiation in this disease stage. Hence, *EZH2* is unlikely to be a driver of NE differentiation in primary tumors. However, its concurrent expression with SPINK1 or TUBB3 in some $\,$ circumstances could be linked to the emergence of NE features. Of therapeutic relevance, many inhibitors directed against AURKA and EZH2 has been developed these recent years [55-57]. Thus, if this hypothesis is confirmed, this could provide a biological rationale for testing the effect of such new-targeted therapies to treat these PCa subgroups. In addition, because EZH2 and/or AURKA upregulation might represent two key events during the transformation of prostate adenocarcinoma towards Neuroendocrine PCa [42,43,58], one could consider targeting these components at an early stage in order to prevent NEPC development and its progression [43,59].

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